

Literature Review**UPDATE ON MANAGEMENT OF DURAL ARTERIOVENOUS FISTULAS**

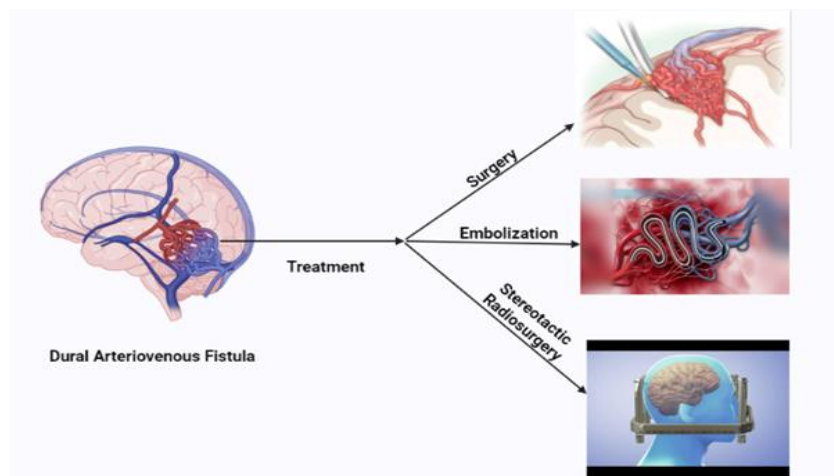
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Abstract: Dural Arteriovenous Fistulas (AVF) represent about 10% of all intracranial vascular lesions. Although they seem benign in nature, the presence of retrograde venous makes them aggressive, with a high risk of complications. Patients may be clinically asymptomatic or experience symptoms ranging from mild to severe hemorrhage, depending on their location. Different treatments are available, but recently, the development of catheter intervention allows most patients to be cured with transcatheter embolization. Stereotactic radiosurgery achieves excellent rates of obliteration for low-grade lesions. In this review, we try to highlight the recent advances in the management of dural AVF.



Keywords: Dural Arteriovenous Fistulas, Embolization, Surgical Ligation, Pathophysiology

INTRODUCTION Dural Arteriovenous Fistulae Dural arteriovenous fistulas (dAVF) are pathologic shunts between arteries and dural veins. They are the most common spinal vascular malformation, representing approximately 10-15% of all intracranial vascular malformations [1-5]. The location of these vascular anomalies is important for risk stratification and prognostication. Spinal dAVF tend to develop in the

thoracic spine, while cranial dAVF are most prevalent in the regions of the transverse, sigmoid, and cavernous sinuses [6-9]. Petroclival dAVF are a rarer but important archetype due to their aggressive nature [6]. Tentorial, galenic, and foramen magnum dAVF are among the most complex and highest-risk lesions [4, 10-12].

There is no clear sex predilection, and there is heterogeneity within the age of presentation, but patients commonly present in the 5th and 6th decades of life [13-18]. There is debate surrounding the formation mechanism; however, it is universally accepted that most

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of these lesions develop idiopathically [19-21]. Herein lies the challenge of detecting these lesions as they are without a true, primary, identifiable cause. Often, they are incidental findings or present symptomatically and further along their clinical course.

Dural venous sinus thrombosis or progressive stenosis is the proposed most likely cause of dAVF. Therefore, it is well documented that hypercoagulable states may predispose individuals to dAVF formation. This is supported by the correlation between prothrombotic conditions such as factor V Leiden, protein C, and protein S deficiency and greater frequency of dAVF [19, 22]. Additional causes of hypercoagulation, including hormonal alterations, pregnancy, and menopause, have also been implicated [3]. Other risk factors for developing dAVF include a history of traumatic head injury, previous craniotomy, concomitant tumor, or infection [3, 5, 13, 19, 22].

The underlying pathophysiology of dAVF is occlusion from inflammation, thromboses, and stenosis causing congestion and venous hypertension. The subsequent elevation in venous pressure results in the formation of fistulous connections between meningeal arteries, dural sinuses, and cortical veins as a compensatory mechanism [19, 21]. Over time, abnormal flow can pivot and become retrograde, leading to cortical venous reflux (CVR); this is the hallmark sign of an aggressive dAVF and is associated with increased morbidity [3, 12, 19]. Due to their progressive nature, dAVF can present symptomatically unless found incidentally. Symptoms are non-specific and depend on the fistula location and the venous drainage pattern. They are classified as high-grade or low-grade, which will be discussed in further detail.

Briefly, low-grade fistulas are generally benign but can present with headache, pulsatile tinnitus, vertigo, bruits, and ophthalmologic abnormalities and visual changes [21, 23-25]. Symptom severity is variable and can be minor or severely affect the quality of life. Clinical symptoms can also vary by location; for instance, dAVF at the cavernous sinus frequently presents with ocular symptoms, including ophthalmoplegia, proptosis, chemosis, and orbital pain [22, 26]. Ocular manifestations may become so apparent late in the course of carotid dAVF that the clinical examination alone can suggest a possible diagnosis [27]. High-grade dAVF are defined by the presence of cortical venous drainage (CVD) and are associated with significant mortality and morbidity. They are aggressive lesions with elevated risks of intracranial hemorrhage (ICH) and non-

hemorrhagic neurological deficits (NHND) [13, 20, 23, 28-30]. In high-grade dAVF, the risk of ICH is reportedly as high as 20% [13]. Furthermore, if left untreated, the risk of rebleeding after ICH in a ruptured dAVF is 35% within the first two weeks [13, 20, 31, 32]. NHND includes seizures, focal nerve deficits, mental status changes, and progressive myelopathy; symptoms will vary with location [3, 19, 24, 27]. Overall, the combined risk of a severe adverse neurological event is approximately 15% annually, with a mortality rate greater than 10% [22, 27, 28, 31]. For this reason, a full work-up and evaluation are warranted if there is clinical suspicion of a dural fistulous lesion. Prompt treatment is recommended for high-grade dAVF to achieve complete obliteration of the fistula.

Several grading scales have been developed to classify dAVF as low- or high-grade. They are based on the venous outflow and architecture and are used to guide treatment recommendations. Historically, the Borden and Cognard classifications have offered the greatest utility and are the most universally referenced [27, 33]. Borden and colleagues [34] designed a three-category classification scheme to describe dAVF: Type I drains directly into a dural venous sinus or meningeal vein, Type II drains into the venous sinus with retrograde drainage into subarachnoid veins, and type III drains directly into subarachnoid veins. In this system, Type II and Type III dAVF are considered high-grade due to retrograde flow and/or cortical venous drainage, which confer a worse natural history.

Similarly, the Cognard classification delineates dAVF by the presence or absence of retrograde flow and CVR, however, it is an eight-category scale. Types I – Type V, with three subsets of Type II make up this classification scheme [35] (Table 1), (Figure 1). Type I and Type IIa are considered low-grade and equivalent to Borden Type I dAVF with drainage directly into the dural sinus. Type IIa differs by the presence of retrograde flow. Type IIb-Type V are high-grade dAVF exhibiting direct cortical venous drainage, retrograde flow, venous ectasia, and drainage into the perimedullary veins [5, 19, 35].

The Zipfel classification, developed by Zipfel and colleagues [36], registers the presence or absence of aggressive symptoms to subcategorize high-grade dAVF further. The rationale for assessing the lesions based on clinical presentation is that mortality and morbidity are significantly higher when CVD is present with associated ICH or NHND. Zipfel Type I dAVF solely drains into dural sinuses, analogous to Borden Type I and Cognard Types I and IIa lesions. Types II and III lesions display CVR and direct cortical venous drainage, respectively.

Borden			Cognard			Zipfel		
Type	Venous Drainage	CVR	Type	Venous Drainage	CVR	Type	Venous Drainage	CVR
Type 1	Dural sinus antegrade	-	Type 1	Dural sinus antegrade	-	Type 1	Dural sinus	-
Type 2	Dural sinus retrograde	+	Type 2a	Dural sinus retrograde	-	Type 2a	Dural sinus	+/-, asymptomatic
Type 3	Cortical vein	+	Type 2b	Dural sinus antegrade	+	Type 2s	Dural sinus	+/-, symptomatic
			Type 2a+b	Dural sinus retrograde	+	Type 3a	Cortical vein	+, asymptomatic
			Type 3	Cortical vein	+	Type 3s	Cortical vein	+, symptomatic
			Type 4	Cortical vein	+ w/ venous ectasia			
			Type 5	Perimedullary veins	+			
Lasjaunias-Geibprasert			Lawton					
Type	Venous Drainage	Type	Venous Drainage	Location	Dural Base	Venous Sinus		
Ventral	Ventral venous plexuses	Type 1, Galenic	Supra/infratentorial	Midline	Anterior falcotentorial junction	Vein of Galen		
Dorsal	Dural venous sinuses	Type 2, Straight Sinus	Infratentorial	Midline	Middle falcotentorial junction	Straight Sinus		
Lateral	Cortical veins	Type 3, Torcular	Supratentorial	Midline	Posterior falcotentorial junction	Torcula		
		Type 4, Tentorial Sinus	Infratentorial	Paramedian	Tentorium	Tentorial Sinus		
		Type 5, Superior Petrosal Sinus	Supratentorial	Lateral	Petrotentorial junction	Superior Petrosal Sinus		
		Type 6, Incisural	Infratentorial	Paramedian	Tentorial incisura	None		

Table 1. Dural arteriovenous fistula classification.

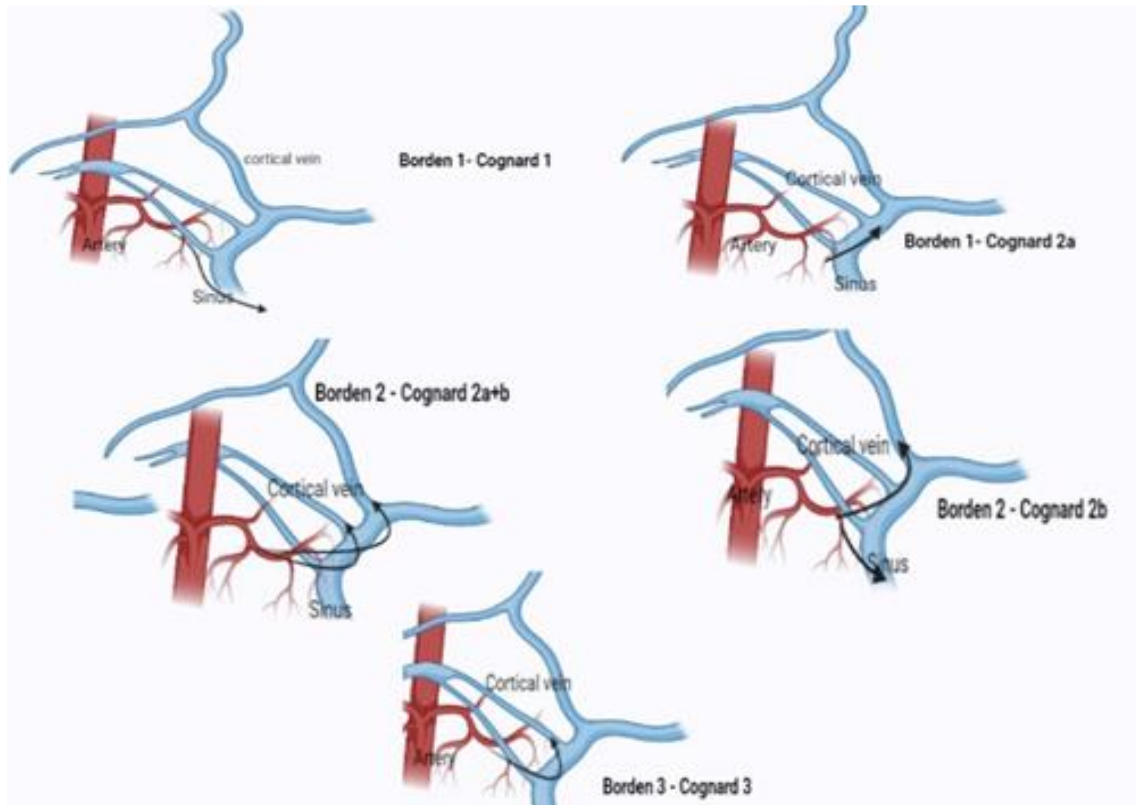


Figure 1. Dural arteriovenous fistula classification.

They are further subdivided into Type II/III A and S, corresponding to asymptomatic and symptomatic presentation [5, 19]. This modification improves accuracy for stratifying which high-grade lesions are of urgent concern versus emergent.

Lasjaunias-Geibprasert and colleagues determined that cranial and spinal dAVFs could be categorized into lesions of ventral, dorsal, and lateral origin [2, 13]. The notion that the embryological origin of the fistula could further illustrate risk is the impetus for developing their system. Accordingly, women are predisposed to ventral origin lesions that drain towards venous plexuses and are typically benign. Dorsal origin fistulas are present at the transverse and sigmoid sinus, draining into dural venous sinuses. The lateral lesions constitute those at the petrous, ethmoid, and tentorial regions and are more common in men. Lateral lesions are also the most frequently discovered SDAVF, and they have the greatest likelihood of being aggressive due to direct CVD and drainage to spinal perimedullary veins, which can cause debilitating myelopathy [8, 13, 37]. Lawton et al. created a new classification scheme for tentorial dAVFs based on fistula

location, dural base, the associated venous sinus, and the direction of venous drainage. This classification system, made up of 6 different subtypes, produces an algorithmic approach to the surgical management of tentorial dAVF to achieve better treatment outcomes. Tentorial dAVFs have a high risk of hemorrhage and require microsurgical interruption of the draining veins because they often cannot be obliterated endovascularly [38].

Additional, more nuanced classifications to label specific types of dAVF have also been produced. The existence of a unique dAVF that has features of Cognard Type IV and Type V lesions, aptly named Type IV + V, has been reported [20]. Though rare, the presence of CVD and venous ectasia with pial medullary drainage is concerning and is an indicator for urgent treatment, even in an asymptomatic presentation.

The DES scheme was developed to examine the inherent heterogeneity among high-grade dAVF. It aims to precisely describe the anatomical localization of the shunt and the characteristics of leptomeningeal venous reflux [39]. Shunts can be composed of bridging veins, dural sinuses,

emissary veins, or exist as isolated sinus shunts. If present, leptomeningeal reflux is subdivided into direct, exclusive, or strained. Using the DES scheme, Baltasavias et al. [39] discovered that lesions with the most aggressive presentation had a bridging vein shunt with strained leptomeningeal reflux. These dAVFs were more amenable to microsurgery than endovascular therapy. Their results illustrate the pertinence of accurately ascribing the true risk of dAVF. It can influence when and what kind of intervention is necessary.

CVR is the hallmark sign of high-grade dAVF and is significantly associated with worse mortality and morbidity. The most consequential symptoms are NHND and ICH due to rupture [40, 41]; however, these are not the most common symptoms upon presentation. Patients with both low- and high-grade dAVF can present asymptotically or most frequently with headache, followed by pulsatile tinnitus [23]. The low-grade dAVF are often considered benign, and treatment is conservative or reserved for symptom management to improve quality of life. There are instances of conversion from low-grade to high-grade, so patients with a known history should be followed for changes in symptoms and monitored with serial imaging [13, 19, 28, 42].

Because dAVFs do not present with a pathognomonic clinical picture, the initial evaluation is with CT and MRI. CT has limited utility and poor sensitivity and specificity for detecting dAVFs. It can detect bleeding, vasogenic edema, and the presence of cortical venous hypertension [19]. MRI is superior to CT, given its better resolution, and some MRI series, such as post-contrast T2, may show leptomeningeal and medullary vessel dilatation, venous ectasia, or venous sinus thrombosis [19, 20]. Hyperintensity on T2/FLAIR sequencing is an important indicator for aggressive dAVFs. Robust FLAIR signaling was identified in patients with CVD presenting with ICH and NHND [43]. Additionally, arterial spin labeling (ASL) reportedly has a high specificity and sensitivity to identify dAVF [44, 45]. The utility of CT and MRI is limited, and they are best used as initial imaging modalities to support clinical suspicions. The gold standard for diagnosis remains diagnostic subtraction angiography (DSA) [20, 44, 46, 47]. Though it is an invasive procedure, DSA is necessary for visualizing arterial feeders, defining the venous architecture, and planning treatment strategies and approaches [3, 13, 19, 20, 31]. The current classification scores used to guide treatment and identify the presence of CVD, venous ectasia, etc., rely on DSA.

The presence of CVD and aggressive symptoms is an absolute indication of treatment. The treatment should be aimed at complete obliteration of the fistula to protect against future hemorrhage and NHND [10, 13, 20]. The presence of asymptomatic CVD requires a more nuanced approach, as these lesions have the potential to progress. A watch-and-wait approach with serial imaging or obliteration on an elective basis may be warranted; this tends to be case-dependent [27]. Currently, the first-line therapy for dAVF is endovascular embolization [16, 48]; however, there are instances where microsurgery, or combined therapy, is more suitable [49]. Surgical obliteration is often more favorable for complex lesions along the tentorium, in the anterior cranial fossa, and involving the transverse and sigmoid sinuses [50]. Endovascular therapy has evolved with advancements in techniques and embolic agents resulting in satisfactory treatment outcomes, but the fistula's architecture limits its efficacy. A proper approach via transarterial or transvenous access is lesion-dependent [30]. The benefit of surgery, though more invasive, is the near 100% obliteration rate and angiographic cure that is routinely achieved [51].

Stereotactic radiosurgery (SRS) is an alternative option usually reserved as a follow-up treatment or for lesions not amenable to surgical or endovascular therapy [31]. SRS is minimally invasive and effective, but dAVFs obliteration rates are reportedly suboptimal [24]. Moreover, there is a latency period between the date of SRS treatment and when the therapy takes effect. It is a poor treatment modality for high-risk lesions that have caused ICH or are at risk of bleeding [28], but it may be a suitable option for appropriately selected low-grade dAVF [31].

Management strategies in cranial dAVF Cranial dural arteriovenous fistulae (dAVF) are vascular malformations consisting of a nidus of arteriovenous shunting within the dura mater. The pathogenesis of this condition involves numerous etiologies, though trauma, previous surgery, venous stenosis, or sinus thrombosis are common causes [31]. While most often benign, retrograde venous drainage and cortical venous reflux cause these structures to carry a high risk of catastrophic intracerebral hemorrhage [31, 52, 53]. Thus, managing cranial dAVF requires careful evaluation to optimize patient outcomes (Figure 2).

Treatment strategies involve the assessment of individual patient characteristics, symptoms, and the risk of adverse events [19]. Of special consideration during dAVF workup is the malformation's anatomic location and venous

drainage pattern, as these factors carry the greatest import on patient symptoms and hemorrhage risk [54]. A conservative approach with regular imaging is preferred if a dAVF is determined to be sufficiently benign (Grades I and II) [54]. Another non-invasive, low-cost treatment that has been successful in a minority of patients, especially those without retrograde venous drainage or severely declined visual acuity, is carotid artery manual compression [55-58]. When the clinical picture demonstrates a more aggressive dAVF, treatment is generally necessary via either microsurgery, transvenous embolization, or stereotactic radiosurgery [59-61].

insufficient or have previously failed, microsurgical disconnection may be used in isolation or tandem with endovascular treatments [68, 69]. While microsurgery is generally safe and highly effective, open surgery brings increased complication risk; thus, this modality is commonly reserved for dAVF, which cannot be treated otherwise or in acute dAVF hemorrhage [34, 70-73].

Lastly, stereotactic radiosurgery is an avenue whereby less aggressive dAVF might be addressed [74]. Radiosurgery gradually obliterates the lesion, which can help prevent venous hypertension or infarction [75]. However, this

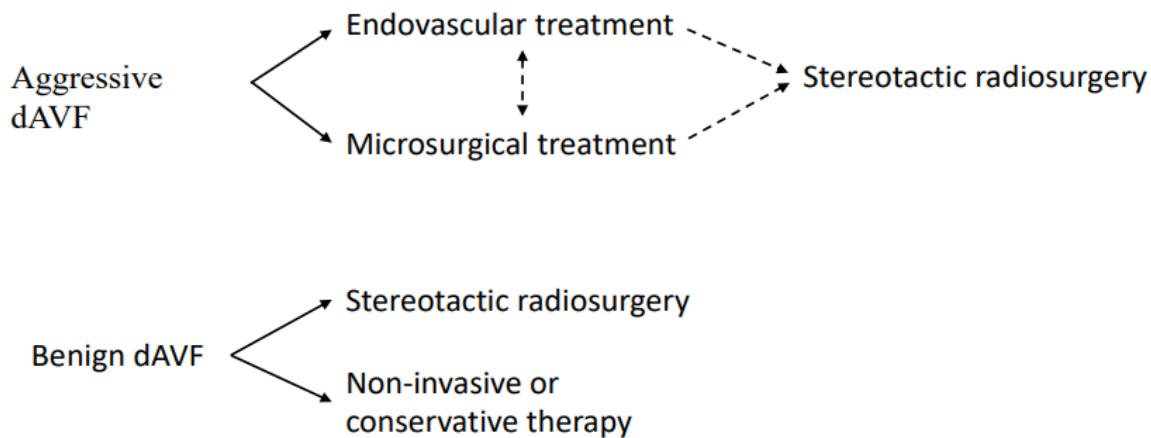


Figure 2. Flowchart outlining the treatment modalities for either benign or aggressive cranial dAVFs. Aggressive dAVF are standardly addressed endovascularly, though microsurgery may be indicated in rare instances. Endovascular and microsurgical approaches may be combined in select dAVF circumstances. If total lesion obliteration is not achieved via endovascular or microsurgical approaches, stereotactic radiosurgery may be employed. For benign lesions, conservative therapy is almost always preferred, though radiosurgery may help reduce patient symptoms.

During invasive treatment of dAVF, complete obliteration of the fistula is ideal, as any residual lesion could lead to dAVF recurrence [31]. In the modern era, endovascular embolization has become the preferred cranial dAVF treatment modality with either an arterial or venous approach [19, 22, 62-65]. Using specialized coils, embolic agents, or particulates, the dAVF may be partially or fully occluded [6, 66, 67]. Careful attention must be given before embolization to fully appreciate the anatomical relationships of the dAVF, including the neurological structures supplied by the dAVF feeder arteries, the presence of anastomoses, and dAVF contributions to nervous tissues [58]. When endovascular approaches are

delayed fistula closure can allow time for adverse events in the presence of aggressive dAVF [76]. For this reason, aggressive dAVF are recommended for endovascular or microsurgical treatments first, with radiosurgery being utilized as secondary therapy to manage residual components [75]. Due to radiosurgery's comparatively less-invasive nature, complications related to this procedure are rare, and rates of dAVF obliteration via radiosurgery are high. Thus, the radiosurgical approach has a firm place in the management of cranial dAVF.

Embolization of dAVF (Figure 3); dAVF can be treated in several ways, the most common of which is endovascular

embolization. Though endovascular embolization is the primary treatment method for most dAVF, the correct technique should be chosen according to the overall presentation, patient age, comorbidities, and prior medical history of hemorrhage [31, 77, 78]. Other

CVR resulted in lower intracranial hemorrhage rates (5.9% vs. 18.2%) and lower non-hemorrhagic neurological deficit rates (0% vs. 27.3%) than symptomatic CVR cases, [82]; however, obliteration is still recommended in these cases [78]. Although dAVFs are commonly treated with

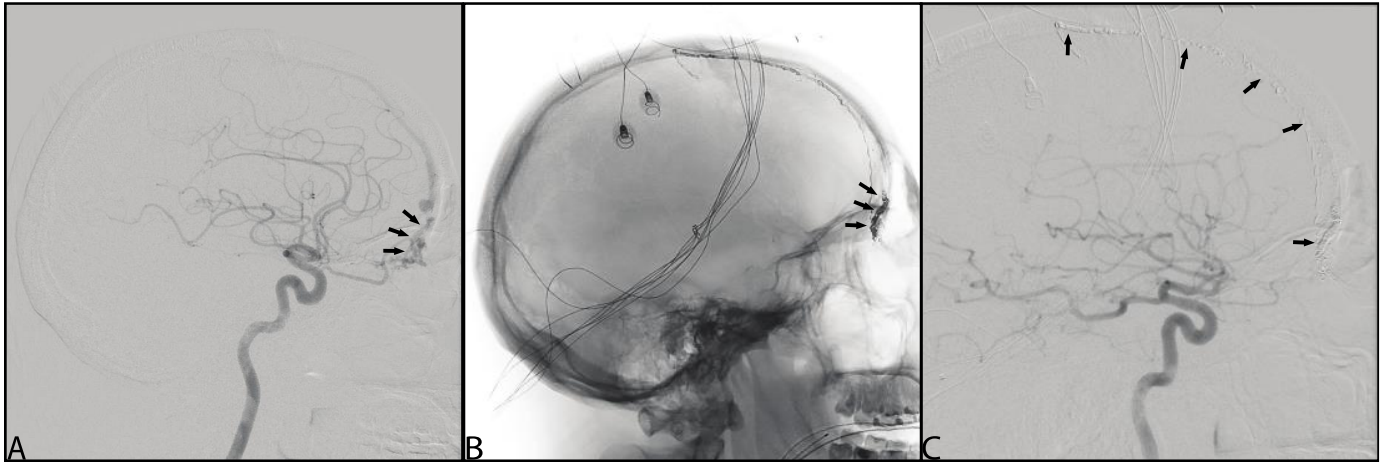


Figure 3. Digital subtraction angiography scans illustrating the embolic agents Onyx and microcoils for the embolization of a cerebral ethmoidal dAVF. A) Right anterior ethmoidal artery angiogram before embolization, arrows point to the dAVF; B) Right anterior ethmoidal artery angiogram after embolization with coils. Arrows point to the microcoils; C) Right anterior ethmoidal artery angiogram after transcranial embolization with Onyx.

important considerations for endovascular management include hemodynamics, angiographic characteristics, and location [76-78]. Alternatively, in cases without cortical venous reflux (CVR) and venous ectasia, dAVF can be managed conservatively [76, 77, 79, 80]. The choice of embolization can also be influenced by hemorrhagic risks, which may, in part, be ascertained from the Borden or Cognard type [35, 81]. The literature suggests that dAVF of Borden Type 2 and 3 (dAVF with CVR) should be treated rapidly due to a higher incidence of intracranial hemorrhage and non-hemorrhagic neurological deficits [79, 82, 83]. Jung Tae Oh et al. report hemorrhage and aggressive symptom rates according to a Borden classification based on a retrospective analysis of 95 patients [81]. In this study, Type I dAVF was associated with a 3% risk of hemorrhage, Type II was associated with a 17% risk of hemorrhage, and Type III had an associated risk of hemorrhage of 46%. Interestingly, in this same study, the authors report success rates (complete obliteration) based on intervention type. The treatment choice of "embolization alone" led to complete obliteration in 80% of cases [81]. In a different study, Strom et al. found that asymptomatic cases of dAVF with

endovascular embolization techniques, they can also be treated using stereotactic radiosurgery when indicated, as is the case for cavernous dAVF without cortical venous drainage or with surgery, which is frequently employed for ethmoidal dAVF [84-87].

Embolization for dAVF is performed transarterial, transvenous, a combination of the two, or through direct percutaneous approaches [78]. Transarterial embolization is performed by placing a microcatheter into an artery in the leg or arm and leading it to the brain through the ophthalmic, ethmoidal, or middle meningeal artery, amongst others [88-90]. For a transvenous approach, the catheter is inserted into the femoral vein and advanced to the brain using intraoperative cone beam CT in much the same way as the transarterial approach [91-93]. Direct percutaneous embolization is often employed for dAVF of the anterior cranial fossa, hypoglossal canal, or upon failure or prior contraindication of a transarterial or transvenous approach [94-98]. The customary practice involves percutaneous or transorbital needle puncture or direct surgical access to expose the vessel, followed by catheterization [78, 94, 99-102]. Several variant

procedures exist using balloon-assisted techniques. Jagadeesan et al. report a technique in which a balloon is inflated proximal to the microcatheter tip at the nidus of the dAVF to negate the need to form an onyx plug [103]. When navigating from larger parent vessels into smaller or tortuous vessels, a common problem occurs where the catheter cannot successfully navigate the bend and transition into the distal vessel. Zhao et al. report a technique where a balloon is inflated to enlarge the parent vessel's lumen, thereby facilitating the microcatheter's escort into the smaller, distal vessels, on the way to the nidus and prior to embolization [104]. The transarterial approach is most frequently indicated of all embolization techniques due to a lower incidence of intra- and postoperative complications and a higher frequency of success [78, 105].

The primary embolic agent used is "Onyx," a non-adhesive polymer made of ethylene vinyl alcohol, dimethyl sulfoxide (DMSO), and micronized tantalum powder (Figure 1) [67, 78, 106-108]. The use of Onyx first appeared in the literature in 1990 and 1991 [109, 110]. As a non-adhesive, one benefit of Onyx is that it doesn't adhere to the microcatheter tip to the extent of traditional acrylic glues. Before Onyx, this limitation complicated retrieval of the microcatheter following the obliteration of the dAVF [111, 112]. One drawback to Onyx is the need for DMSO during its administration which can cause chemical irritation to the endothelium or, in rare cases, may result in vasospasms [113-115]. One alternative is the embolic agent n-butyl-2-cyanoacrylate, commonly known as acrylic glue, although its use has decreased with the advent of Onyx [116-119]. One of the beneficial features of acrylic glue is its quick preparation and administration capabilities [112, 120, 121]. This can play an especially important role in emergent cases. Acrylic glue is cheaper than Onyx, making it more accessible for low-income countries [117]. However, one drawback of acrylic glue is its rapid polymerization following contact with ionic solutions [122].

Additionally, unlike Onyx, acrylic glue is not radiopaque, which can make intraoperative visualization more laborious. Coils are another embolic agent (Figure 1). Coils are typically made of steel or platinum and can be created according to different specifications. Coils are frequently combined with Onyx to achieve complete obliteration [123-125]. Risks include coil migration and subsequent obstruction of adjacent lumina [126]. The advantages of coils include quick preparation, administration, and greater visualization. Some coils come with coatings like

hydrogel, which increases the diameter of coils upon contact with blood [127]. Many coils are also surrounded by wool or other thrombogenic agents. Of all the embolic agents available, Onyx has quickly expanded to become the primary choice for dAVFs, with reported complete obliteration rates between 55 and 60 percent [128, 129]. The odds of avoiding surgery following embolization with Onyx are also significantly higher at 81.80 percent compared to 22.22 percent with acrylic glue [130].

Surgical management of dAVF The anatomical location of the fistula and its effect on the venous drainage flow dynamics produces symptoms that require treatment. The presenting clinical features are variable and depend on the location of the fistula and include seizures, myelopathy, cranial nerve palsies, and sensory or motor deficits. In about 20%–33% of dAVFs, the presentation is intracranial hemorrhage [131]. Management lines include both endovascular and surgical approaches. While the endovascular approach is widely used now, surgery is still a safe available option [70]. Over 80 years, the technique of surgery for AVM has progressed steadily since the first resection. Introducing cerebral angiography in 1927 by Antonio Caetano de Abreu Freire Egas Moniz has accelerated the surgical progress [132]. Surgical treatment is always considered for all aggressive dural AVF [133]. In most cases of cranial dural AVF involving the transverse sinus, the AVF is drained by the sinus itself or the dural veins. These types of AVF tend to develop new collateral venous routes with the sinus or dural veins. Therefore, surgical excision is considered the definitive therapy in these cases [134]. Other indications for surgery include abnormal tortuous arterial feeders not suitable for embolization, feeders involved in normal brain structures feeding or difficult to localize or a fistula that persists after endovascular or radiosurgery [135].

Surgery is always considered for high-grade dural AVF with retrograde cortical lesions. These lesions have a high rate of complications than the lower grade [136]. Surgical-wise, dural AVFs are classified into two types direct and indirect. The direct types usually have a specific arterial feeder to a draining vein, while the indirect types have numerous small arterial feeders that pass through the dura [135]. Most of the surgical techniques for dural AVM are like those involved in the treatment of dural AVF except for the nidus which is a feature of dural AVM only [68]. In 1974, Hugosson and Bergstrom described sinus skeletonization for the indirect type [137]. Lucas et al also described the same technique in two cases of dural AVF involving the tentorial apex [138]. While sinus skeletonization preserves

the patency of the sinus, it still has some burdens. Brain retraction, air embolism, and bleeding are possible identified common complications [139].

Spinal Dural AVF (Figure 4); Spinal dAVFs are direct communication between a radiculomeningeal artery and a radicular vein [140-143]. They represent only 1-2% of

can be managed successfully in the majority of patients if the diagnosis is made before irreversible neurological deficits develop [159]. Available options include surgical disconnection, endovascular embolization, or a combination of the two. Surgery is largely pursued due to the simplicity of the procedure, as well as the low morbidity and recurrence rates observed [150]. Recently,

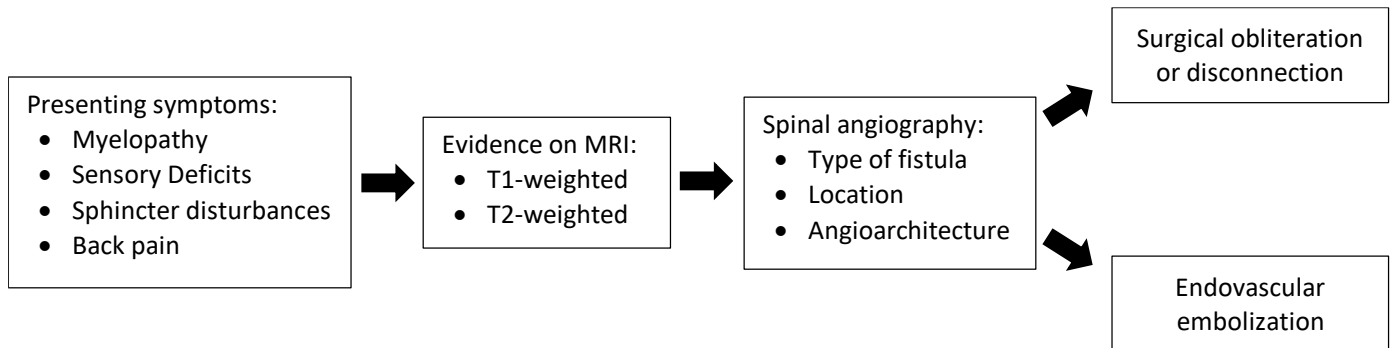


Figure 4. Patient Course of Spinal dAVF. Patient presents with classic symptoms and undergoes an MRI that suggests spinal dAVF. MRI findings prompt spinal angiography, the gold standard diagnostic tool for spinal dAVF. Based on findings from the angiography, as well as evidence based and shared decision making between the provider and patient.

vascular neurologic pathologies, though are the most common spinal vascular malformation [142, 144-147]. Spinal dAVFs occur primarily in the thoraco-lumbar region, and rarely in the cervical region [148-150]. Cases are primarily idiopathic, though may develop following trauma or surgery.[150, 151] Classically, spinal dAVFs present with progressive spastic motor weakness, sensory deficits, sphincter disturbances, and back pain [148, 150-153]. Other patients may present with hemorrhage or an acute exacerbation referred to as “Foix-Alajouanine syndrome.”[152]. Myelopathy, intramedullary edema, and chronic hypoxia develop due to the venous hypertension from arterial blood shunting into the valveless veins of the spinal cord [144, 153-155]. Subsequently, a decrease in arterial supply, arterial steal, and ischemia are observed [153-155]. This myelopathy can be irreversible if left untreated [153-155].

Initially, both T1-weighted and T2-weighted MRIs often provide the first evidence suggestive of a spinal dAVF via a serpentine pattern of low signal in the subarachnoid space [150]. Spinal angiography is the diagnostic gold standard, as these studies precisely distinguish the type of fistula, position of fistula, and its angioarchitecture [156, 157]. From a treatment standpoint, the goal in spinal dAVFs is eliminating the venous congestion and giving the spinal cord an environment to recover [154, 158]. Spinal dAVFs

advances in endovascular techniques have drastically increased the number of patients pursuing embolization options[147, 148, 152, 154, 160], though literature seems to continue to favor primary surgical treatment from failure rate and late recurrence standpoints [161, 162]. Microscopic direct surgery is considered in cases when endovascular access is not safely possible [163].

Previous meta-analysis by Steinmetz and colleagues found improvements of symptoms to favor surgical management [160]. In 2021, Fiaschi and colleagues found improvement of motor symptoms in >80% of patients following surgical obliteration and improvement of sphincter dysfunction in >30%. Additionally, their cohort demonstrated higher patient quality-of-life perceptions following surgical management when compared to embolization [164].

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REFERENCES

- Cannizzaro, D., et al., Changing Clinical and Therapeutic Trends in Tentorial Dural Arteriovenous Fistulas: A Systematic Review. *AJNR Am J Neuroradiol*, 2015. 36(10): p. 1905-11.
- Cannizzaro, D., et al., Endovascular and surgical approaches of ethmoidal dural fistulas: a multicenter experience and a literature review. *Neurosurg Rev*, 2018. 41(2): p. 391-398.
- Chaichana, K.L., et al., Dural arteriovenous fistulas: epidemiology and clinical presentation. *Neurosurg Clin N Am*, 2012. 23(1): p. 7-13.
- Della Pepa, G.M., et al., Angio-Architectural Features of High-Grade Intracranial Dural Arteriovenous Fistulas: Correlation With Aggressive Clinical Presentation and Hemorrhagic Risk. *Neurosurgery*, 2017. 81(2): p. 315-330.
- Gomez, J., et al., Classification schemes of cranial dural arteriovenous fistulas. *Neurosurg Clin N Am*, 2012. 23(1): p. 55-62.
- Hou, K., et al., Endovascular treatment for dural arteriovenous fistulas in the petroclival region. *Int J Med Sci*, 2020. 17(18): p. 3020-3030.
- Hou, K., et al., Endovascular treatment of the cavernous sinus dural arteriovenous fistula: current status and considerations. *Int J Med Sci*, 2020. 17(8): p. 1121-1130.
- Krings, T., Vascular malformations of the spine and spinal cord* : anatomy, classification, treatment. *Clin Neuroradiol*, 2010. 20(1): p. 5-24.
- Soo Kim, M., Clinical characteristics of dural arteriovenous fistula, D.H. Han, et al., Editors. 2002, *J Clin Neurosci*. p. 147-55.
- Caton, M.T., et al., Dural Arteriovenous Fistulas of the Foramen Magnum Region: Clinical Features and Angioarchitectural Phenotypes. *AJNR Am J Neuroradiol*, 2021. 42(8): p. 1486-1491.
- Cohen, J.E., et al., Clinical and angioarchitectural factors influencing the endovascular approach to galenic dural arteriovenous fistulas in adults: case series and review of the literature. *Acta Neurochir (Wien)*, 2017. 159(5): p. 845-853.
- Signorelli, F., et al., Diagnosis and management of dural arteriovenous fistulas: a 10 years single-center experience. *Clin Neurol Neurosurg*, 2015. 128: p. 123-9.
- Bhatia, K.D., et al., Endovascular Management of Intracranial Dural AVFs: Principles. *AJNR Am J Neuroradiol*, 2022. 43(2): p. 160-166.
- Maus, V., et al., Endovascular Treatment of Intracranial Dural Arteriovenous Fistulas: A German Single-Center Experience. *Cerebrovasc Dis Extra*, 2020. 10(2): p. 84-93.
- Negro, A., et al., Intracranial Hemorrhage from Dural Arteriovenous Fistulas: What Can We Find with CT Angiography? *Tomography*, 2021. 7(4): p. 804-814.
- Qureshi, A.M., et al., Clinical and Angioarchitectural Features of Ruptured Dural Arteriovenous Fistulas. *World Neurosurg*, 2021. 147: p. e476-e481.
- Samaniego, E.A., et al., Dural arteriovenous fistulas without cortical venous drainage: presentation, treatment, and outcomes. *J Neurosurg*, 2022. 136(4): p. 942-950.
- Xu, K., et al., Current status of endovascular treatment for dural arteriovenous fistulae in the anterior cranial fossa: A systematic literature review. *Int J Med Sci*, 2019. 16(2): p. 203-211.
- Reynolds, M.R., G. Lanzino, and G.J. Zipfel, Intracranial Dural Arteriovenous Fistulae. *Stroke*, 2017. 48(5): p. 1424-1431.
- Casasco, A., et al., A new subtype of intracranial dural AVF according to the patterns of venous drainage. *Interv Neuroradiol*, 2021. 27(1): p. 121-128.
- Corbelli, I., et al., Dural arteriovenous fistulas and headache features: an observational study. *J Headache Pain*, 2020. 21(1): p. 6.
- Gandhi, D., et al., Intracranial dural arteriovenous fistulas: classification, imaging findings, and treatment. *AJNR Am J Neuroradiol*, 2012. 33(6): p. 1007-13.
- Haas, L.J., et al., Prevalence of Tinnitus in Patients Diagnosed with Cerebral Arteriovenous Fistula Treated with Endovascular Technique. *Int Arch Otorhinolaryngol*, 2022. 26(3): p. e428-e433.
- Hasegawa, H., et al., A Practical Grading Scale for Predicting Outcomes of Radiosurgery for Dural Arteriovenous Fistulas: JLGK 1802 Study. *J Stroke*, 2022. 24(2): p. 278-287.

25. Satomi, J., et al., Benign cranial dural arteriovenous fistulas: outcome of conservative management based on the natural history of the lesion. *J Neurosurg*, 2002. 97(4): p. 767-70.
26. Pan, D.H., et al., Intracranial dural arteriovenous fistulas: natural history and rationale for treatment with stereotactic radiosurgery. *Prog Neurol Surg*, 2013. 27: p. 176-94.
27. Colby, G.P., et al., Historical perspective of treatments of cranial arteriovenous malformations and dural arteriovenous fistulas. *Neurosurg Clin N Am*, 2012. 23(1): p. 15-25.
28. Chen, C.J., et al., Intervention for unruptured high-grade intracranial dural arteriovenous fistulas: a multicenter study. *J Neurosurg*, 2022. 136(4): p. 962-970.
29. Kuwayama, N., Epidemiologic Survey of Dural Arteriovenous Fistulas in Japan: Clinical Frequency and Present Status of Treatment. *Acta Neurochir Suppl*, 2016. 123: p. 185-8.
30. Li, C., et al., Clinical and Angioarchitectural Risk Factors Associated with Intracranial Hemorrhage in Dural Arteriovenous Fistulas: A Single-Center Retrospective Study. *PLoS One*, 2015. 10(6): p. e0131235.
31. Baharvahdat, H., et al., Updates in the management of cranial dural arteriovenous fistula. *Stroke Vasc Neurol*, 2020. 5(1): p. 50-58.
32. Gross, B.A., et al., Evolution of treatment and a detailed analysis of occlusion, recurrence, and clinical outcomes in an endovascular library of 260 dural arteriovenous fistulas. *J Neurosurg*, 2017. 126(6): p. 1884-1893.
33. Hiramatsu, M., et al., Epidemiology of dural arteriovenous fistula in Japan: Analysis of Japanese Registry of Neuroendovascular Therapy (JR-NET2). *Neurol Med Chir (Tokyo)*, 2014. 54(1): p. 63-71.
34. Borden, J.A., J.K. Wu, and W.A. Shucart, A proposed classification for spinal and cranial dural arteriovenous fistulous malformations and implications for treatment. *J Neurosurg*, 1995. 82(2): p. 166-79.
35. Cognard, C., et al., Cerebral dural arteriovenous fistulas: clinical and angiographic correlation with a revised classification of venous drainage. *Radiology*, 1995. 194(3): p. 671-80.
36. Zipfel, G.J., et al., Cranial dural arteriovenous fistulas: modification of angiographic classification scales based on new natural history data. *Neurosurg Focus*, 2009. 26(5): p. E14.
37. Spittau, B., et al., Dural arteriovenous fistulas of the hypoglossal canal: systematic review on imaging anatomy, clinical findings, and endovascular management. *J Neurosurg*, 2015. 122(4): p. 883-903.
38. Lawton, M.T., et al., Tentorial dural arteriovenous fistulae: operative strategies and microsurgical results for six types. *Neurosurgery*, 2008. 62(3 Suppl 1): p. 110-24; discussion 124-5.
39. Baltasvias, G., A. Valavanis, and L. Regli, Cranial dural arteriovenous shunts: selection of the ideal lesion for surgical occlusion according to the classification system. *Acta Neurochir (Wien)*, 2019. 161(9): p. 1775-1781.
40. Abecassis, I.J., et al., Assessing the rate, natural history, and treatment trends of intracranial aneurysms in patients with intracranial dural arteriovenous fistulas: a Consortium for Dural Arteriovenous Fistula Outcomes Research (CONDOR) investigation. *J Neurosurg*, 2022. 136(4): p. 971-980.
41. Wen, H.Y., H.C. Chen, and S.T. Yang, Risk Factors of Aggressive Clinical Presentation in Patients with Angiographically Aggressive Cranial Dural Arteriovenous Fistulas. *J Clin Med*, 2021. 10(24).
42. Shah, M.N., et al., Borden-Shucart Type I dural arteriovenous fistulas: clinical course including risk of conversion to higher-grade fistulas. *J Neurosurg*, 2012. 117(3): p. 539-45.
43. Patel, B., et al., T2-Weighted-Fluid-Attenuated Inversion Recovery Hyperintensity on Magnetic Resonance Imaging Is Associated With Aggressive Symptoms in Patients With Dural Arteriovenous Fistulas. *Stroke*, 2019. 50(9): p. 2565-2567.
44. Amukotuwa, S.A., et al., Arterial Spin-Labeling Improves Detection of Intracranial Dural Arteriovenous Fistulas with MRI. *AJNR Am J Neuroradiol*, 2018. 39(4): p. 669-677.
45. Togao, O., et al., Vessel-Selective 4D-MRA Using Superselective Pseudocontinuous Arterial Spin-Labeling with Keyhole and View-Sharing for Visualizing Intracranial Dural AVFs. *AJNR Am J Neuroradiol*, 2022. 43(3): p. 368-375.
46. Lin, Y.H., et al., Diagnostic accuracy of CTA and MRI/MRA in the evaluation of the cortical venous reflux in the intracranial dural arteriovenous fistula DAVF. *Neuroradiology*, 2018. 60(1): p. 7-15.
47. Raman, A., et al., A Systematic Review Comparing Digital Subtraction Angiogram with Magnetic Resonance Angiogram Studies in Demonstrating the Angioarchitecture of Cerebral Arteriovenous Malformations. *Cureus*, 2022. 14(6): p. e25803.

48. Priola, S.M., et al., Angio-architecture of complex cranial dural arteriovenous fistulas: A single centre retrospective review of treatment modalities and outcomes. *J Clin Neurosci*, 2020. 76: p. 87-99.
49. Paul, A.R., Selection of treatment modalities or observation of dural arteriovenous fistulas, G.P. Colby, et al., Editors. 2012, *Neurosurg Clin N Am*. p. 77-85.
50. Shimada, K., et al., Efficacy of intraarterial indocyanine green videoangiography in surgery for arteriovenous fistula at the craniocervical junction in a hybrid operating room: illustrative cases. *J Neurosurg Case Lessons*, 2022. 3(23): p. CASE22100.
51. Paredes, I., et al., [Intracranial dural arteriovenous fistulae. Experience after 81 cases and literature review]. *Neurocirugia (Astur)*, 2013. 24(4): p. 141-51.
52. Halbach, V.V., et al., Treatment of dural fistulas involving the deep cerebral venous system. *AJNR Am J Neuroradiol*, 1989. 10(2): p. 393-9.
53. Awad, I.A., et al., Intracranial dural arteriovenous malformations: factors predisposing to an aggressive neurological course. *J Neurosurg*, 1990. 72(6): p. 839-50.
54. Kim, M.S., et al., Clinical characteristics of dural arteriovenous fistula. *J Clin Neurosci*, 2002. 9(2): p. 147-55.
55. Halbach, V.V., et al., Dural fistulas involving the cavernous sinus: results of treatment in 30 patients. *Radiology*, 1987. 163(2): p. 437-42.
56. Halbach, V.V., et al., Dural fistulas involving the transverse and sigmoid sinuses: results of treatment in 28 patients. *Radiology*, 1987. 163(2): p. 443-7.
57. Kai, Y., et al., External Manual Carotid Compression is Effective in Patients with Cavernous Sinus Dural Arteriovenous Fistulae. *Interv Neuroradiol*, 2007. 13 Suppl 1(Suppl 1): p. 115-22.
58. Sarma, D. and K. ter Brugge, Management of intracranial dural arteriovenous shunts in adults. *Eur J Radiol*, 2003. 46(3): p. 206-20.
59. Klisch, J., et al., Transvenous treatment of carotid cavernous and dural arteriovenous fistulae: results for 31 patients and review of the literature. *Neurosurgery*, 2003. 53(4): p. 836-56; discussion 856-7.
60. Tu, Y.K., H.M. Liu, and S.C. Hu, Direct surgery of carotid cavernous fistulae and dural arteriovenous malformations of the cavernous sinus. *Neurosurgery*, 1997. 41(4): p. 798-805; discussion 805-6.
61. Pollock, B.E., et al., Stereotactic radiosurgery and particulate embolization for cavernous sinus dural arteriovenous fistulae. *Neurosurgery*, 1999. 45(3): p. 459-66; discussion 466-7.
62. Chung, S.J., et al., Intracranial dural arteriovenous fistulas: analysis of 60 patients. *Cerebrovasc Dis*, 2002. 13(2): p. 79-88.
63. Alleyne, C.H., Jr., Z. Numaguchi, and H.Z. Wang, Transarterial Embolisation of Dural Arteriovenous Fistula Involving an Isolated Segment of the Superior Petrosal Sinus. A Case report. *Interv Neuroradiol* 2000. 6(4): p. 337-41.
64. Borg, N., et al., Transarterial Embolization of Ethmoidal Dural Arteriovenous Fistula: 2-Dimensional Video. *Oper Neurosurg (Hagerstown)*, 2022. 22(6): p. e275.
65. Limbucci, N., et al., Transvenous Embolization of Ethmoidal Dural Arteriovenous Fistulas: Case Series and Review of the Literature. *World Neurosurg*, 2018. 110: p. e786-e793.
66. Oh, J.S., et al., Endovascular Treatment of Dural Arteriovenous Fistulas: Single Center Experience. *J Korean Neurosurg Soc*, 2016. 59(1): p. 17-25.
67. Cognard, C., et al., Endovascular treatment of intracranial dural arteriovenous fistulas with cortical venous drainage: new management using Onyx. *AJNR Am J Neuroradiol*, 2008. 29(2): p. 235-41.
68. Wachter, D., et al., Microsurgery can cure most intracranial dural arteriovenous fistulae of the sinus and non-sinus type. *Neurosurg Rev*, 2011. 34(3): p. 337-45; discussion 345.
69. Takai, K., et al., Microsurgical versus endovascular treatment of spinal epidural arteriovenous fistulas with intradural venous drainage: a multicenter study of 81 patients. *J Neurosurg Spine*, 2020: p. 1-11.
70. Collice, M., et al., Surgical treatment of intracranial dural arteriovenous fistulae: role of venous drainage. *Neurosurgery*, 2000. 47(1): p. 56-66; discussion 66-7.
71. Davies, M.A., et al., The natural history and management of intracranial dural arteriovenous fistulae. Part 2: aggressive lesions. *Interv Neuroradiol*, 1997. 3(4): p. 303-11.
72. Lucas, C.P., et al., Treatment for intracranial dural arteriovenous malformations: a meta-analysis from the English language literature. *Neurosurgery*, 1997. 40(6): p. 1119-30; discussion 1130-2.
73. Sorteberg, W., et al., Endovascular versus surgical treatment of cranial dural arteriovenous fistulas: a single-center 8-year experience. *Acta Neurochir (Wien)*, 2022. 164(1): p. 151-161.

74. Pan, D.H., et al., Stereotactic radiosurgery for the treatment of dural arteriovenous fistulas involving the transverse-sigmoid sinus. *J Neurosurg*, 2002. 96(5): p. 823-9.
75. Yang, H.C., et al., Radiosurgery for Dural Arteriovenous Fistulas. *Prog Neurol Surg*, 2019. 34: p. 248-259.
76. Miller, T.R. and D. Gandhi, Intracranial Dural Arteriovenous Fistulae: Clinical Presentation and Management Strategies. *Stroke*, 2015. 46(7): p. 2017-25.
77. Mulholland, C.B., M.Y.S. Kalani, and F.C. Albuquerque, Endovascular management of intracranial dural arteriovenous fistulas. *Handb Clin Neurol*, 2017. 143: p. 117-123.
78. Rammos, S., C. Bortolotti, and G. Lanzino, Endovascular management of intracranial dural arteriovenous fistulae. *Neurosurg Clin N Am*, 2014. 25(3): p. 539-49.
79. van Dijk, J.M., et al., Clinical course of cranial dural arteriovenous fistulas with long-term persistent cortical venous reflux. *Stroke*, 2002. 33(5): p. 1233-6.
80. Sato, K., et al., Compromise of brain tissue caused by cortical venous reflux of intracranial dural arteriovenous fistulas: assessment with diffusion-weighted magnetic resonance imaging. *Stroke*, 2011. 42(4): p. 998-1003.
81. Oh, J.T., et al., Intracranial dural arteriovenous fistulas: clinical characteristics and management based on location and hemodynamics. *J Cerebrovasc Endovasc Neurosurg*, 2012. 14(3): p. 192-202.
82. Strom, R.G., et al., Cranial dural arteriovenous fistulae: asymptomatic cortical venous drainage portends less aggressive clinical course. *Neurosurgery*, 2009. 64(2): p. 241-7; discussion 247-8.
83. Duffau, H., et al., Early rebleeding from intracranial dural arteriovenous fistulas: report of 20 cases and review of the literature. *J Neurosurg*, 1999. 90(1): p. 78-84.
84. Tonetti, D.A., et al., Stereotactic Radiosurgery for Dural Arteriovenous Fistulas without Cortical Venous Reflux. *World Neurosurg*, 2017. 107: p. 371-375.
85. Neurosurgery, U.o.P., Bradley Gross Discusses Dural Arteriovenous Fistulas. 2021.
86. Gross, B.A., et al., Stereotactic radiosurgery for cerebral dural arteriovenous fistulas. *Neurosurg Focus*, 2012. 32(5): p. E18.
87. Gross, B.A., et al., Clinical and Anatomic Insights From a Series of Ethmoidal Dural Arteriovenous Fistulas at Barrow Neurological Institute. *World Neurosurg*, 2016. 93: p. 94-9.
88. Robert, T., et al., Endovascular treatment of cribriform plate dural arteriovenous fistulas: technical difficulties and complications avoidance. *J Neurointerv Surg*, 2016. 8(9): p. 954-8.
89. Tahon, F., et al., Dural arteriovenous fistula of the anterior fossa treated with the Onyx liquid embolic system and the Sonic microcatheter. *Neuroradiology*, 2008. 50(5): p. 429-32.
90. Piergallini, L., et al., Anterior cranial fossa dural arteriovenous fistula: Transarterial embolization from the ophthalmic artery as first-line treatment. *J Neuroradiol*, 2021. 48(3): p. 207-214.
91. Layton, K.F., M.D. Nelson, and D.F. Kallmes, Transarterial coil embolization of the venous component of aggressive type 4 dural arteriovenous fistulas. *AJNR Am J Neuroradiol*, 2006. 27(4): p. 750-2.
92. Okamura, A., et al., Intraoperative cone-beam computed tomography contributes to avoiding hypoglossal nerve palsy during transvenous embolization for dural arteriovenous fistula of the anterior condylar confluence. *Interv Neuroradiol*, 2016. 22(5): p. 584-9.
93. Crockett, M.T., et al., Transvenous coil embolization with intra-operative cone beam CT assistance in the treatment of hypoglossal canal dural arteriovenous fistulae. *J Neurointerv Surg*, 2019. 11(2): p. 179-183.
94. White, J.B., et al., Transorbital puncture for the treatment of cavernous sinus dural arteriovenous fistulas. *AJNR Am J Neuroradiol*, 2007. 28(7): p. 1415-7.
95. Nerva, J.D., D.K. Hallam, and B.V. Ghodke, Percutaneous transfacial direct embolization of an intraosseous dural arteriovenous fistula. *Neurosurgery*, 2014. 10 Suppl 1: p. E178-82.
96. Padhi, R., et al., Direct Superior Ophthalmic Vein Approach to Treat Anterior Condylar Confluence Dural Arteriovenous Fistula. *Neurointervention*, 2021. 16(3): p. 280-284.
97. Diaz, O.M., et al., Unique percutaneous direct puncture technique for occlusion of a hypoglossal canal dural arteriovenous fistula. *J Neurointerv Surg*, 2018. 10(12): p. 1179-1182.
98. Cavalcanti, D.D., et al., Percutaneous transorbital direct puncture to obliterate a cavernous sinus dural arteriovenous fistula. *J Neurointerv Surg*, 2021. 13(12): p. 1190.

99. Lv, M., et al., Direct percutaneous transorbital puncture under fluoroscopic guidance with a 3D skull reconstruction overlay for embolisation of intraorbital and cavernous sinus dural arteriovenous fistulas. *Interv Neuroradiol*, 2015. 21(3): p. 357-61.
100. Saura, P., et al., Direct transforaminal Onyx embolization of intracranial dural arteriovenous fistulas: technical note and report of five cases. *J Neurointerv Surg*, 2014. 6(7): p. 500-4.
101. Chapot, R., et al., Transcranial puncture through the parietal and mastoid foramina for the treatment of dural fistulas. Report of four cases. *J Neurosurg*, 2007. 106(5): p. 912-5.
102. Houdart, E., et al., Transcranial approach for venous embolization of dural arteriovenous fistulas. *J Neurosurg*, 2002. 97(2): p. 280-6.
103. Jagadeesan, B.D., et al., Endovascular balloon-assisted embolization of intracranial and cervical arteriovenous malformations using dual-lumen coaxial balloon microcatheters and Onyx: initial experience. *Neurosurgery*, 2013. 73(2 Suppl Operative): p. ons238-43; discussion ons243.
104. Zhao, W.Y., et al., Balloon-assisted superselective microcatheterization for transarterial treatment of cranial dural arteriovenous fistulas: technique and results. *Neurosurgery*, 2012. 71(2 Suppl Operative): p. ons269-73; discussion ons273.
105. Li, C., et al., Transarterial treatment with Onyx of Cognard type IV anterior cranial fossa dural arteriovenous fistulas. *J Neurointerv Surg*, 2014. 6(2): p. 115-20.
106. Macdonald, J.H., J.S. Millar, and C.S. Barker, Endovascular treatment of cranial dural arteriovenous fistulae: a single-centre, 14-year experience and the impact of Onyx on local practise. *Neuroradiology*, 2010. 52(5): p. 387-95.
107. Hu, Y.C., et al., Cranial dural arteriovenous fistula: transarterial Onyx embolization experience and technical nuances. *J Neurointerv Surg*, 2011. 3(1): p. 5-13.
108. Zhang, J., et al., Transarterial and transvenous embolization for cavernous sinus dural arteriovenous fistulae. *Interv Neuroradiol*, 2010. 16(3): p. 269-77.
109. Taki, W., et al., A new liquid material for embolization of arteriovenous malformations. *AJNR Am J Neuroradiol*, 1990. 11(1): p. 163-8.
110. Terada, T., et al., Embolization of arteriovenous malformations with peripheral aneurysms using ethylene vinyl alcohol copolymer. Report of three cases. *J Neurosurg*, 1991. 75(4): p. 655-60.
111. Vollherbst, D.F., et al., Glue, Onyx, Squid or PHIL? Liquid Embolic Agents for the Embolization of Cerebral Arteriovenous Malformations and Dural Arteriovenous Fistulas. *Clin Neuroradiol*, 2022. 32(1): p. 25-38.
112. Fumarola EM, I.A., Piacentino F, Carrafiello G., Glue or onyx: A guide to choice – tips and techniques. *Journal of Endovascular Resuscitation and Trauma Management*, 2020.
113. Murayama, Y., et al., Nonadhesive liquid embolic agent for cerebral arteriovenous malformations: preliminary histopathological studies in swine rete mirabile. *Neurosurgery*, 1998. 43(5): p. 1164-75.
114. Jahan, R., et al., Embolization of arteriovenous malformations with Onyx: clinicopathological experience in 23 patients. *Neurosurgery*, 2001. 48(5): p. 984-95; discussion 995-7.
115. Nogueira, R.G., et al., Preliminary experience with onyx embolization for the treatment of intracranial dural arteriovenous fistulas. *AJNR Am J Neuroradiol*, 2008. 29(1): p. 91-7.
116. Wakhloo, A.K., et al., Transvenous n-butyl-cyanoacrylate infusion for complex dural carotid cavernous fistulas: technical considerations and clinical outcome. *AJNR Am J Neuroradiol*, 2005. 26(8): p. 1888-97.
117. Jung, J.Y. and J.Y. Lee, Transvenous injection of n-butyl 2-cyanoacrylate to obliterate the pathologic cavernous sinus as a salvage technique for incompletely obliterated complex cavernous sinus dural arteriovenous fistula after transvenous coil embolization. *J Cerebrovasc Endovasc Neurosurg*, 2021. 23(4): p. 348-353.
118. Li, M.H., et al., Trans-arterial embolisation therapy of dural carotid-cavernous fistulae using low concentration n-butyl-cyanoacrylate. *Acta Neurochir (Wien)*, 2008. 150(11): p. 1149-56; discussion 1156.
119. Radvany, M.G. and L. Gregg, Endovascular treatment of cranial arteriovenous malformations and dural arteriovenous fistulas. *Neurosurg Clin N Am*, 2012. 23(1): p. 123-31.
120. Yonemitsu, T., et al., Evaluation of transcatheter arterial embolization with gelatin sponge particles, microcoils, and n-butyl cyanoacrylate for acute arterial bleeding in a coagulopathic condition. *J Vasc Interv Radiol*, 2009. 20(9): p. 1176-87.

121. Abdulmalak, G., et al., Safety and efficacy of transcatheter embolization with Glubran(®)2 cyanoacrylate glue for acute arterial bleeding: a single-center experience with 104 patients. *Abdom Radiol (NY)*, 2018. 43(3): p. 723-733.
122. Cromwell, L.D. and C.W. Kerber, Modification of cyanoacrylate for therapeutic embolization: preliminary experience. *AJR Am J Roentgenol*, 1979. 132(5): p. 799-801.
123. Abaunza-Camacho, J.F., et al., Direct transcranial coil and Onyx embolization of a dural arteriovenous fistula: Technical note and brief literature review. *J Clin Neurosci*, 2020. 80: p. 232-237.
124. Koutsouras, G.W., et al., Coil and Onyx embolization of a torcular herophili dural arteriovenous fistula in a full-term neonate with advanced heart failure using a transumbilical approach. *J Neurosurg Pediatr*, 2018. 23(1): p. 80-85.
125. Nam, T.K., et al., Feasibility and Effectiveness of Direct Puncture and Onyx Embolization for Transverse Sinus Dural Arteriovenous Fistula. *Yonsei Med J*, 2019. 60(11): p. 1112-1115.
126. Vaidya, S., K.R. Tozer, and J. Chen, An overview of embolic agents. *Semin Intervent Radiol*, 2008. 25(3): p. 204-15.
127. Klurfan, P., et al., Transvenous treatment of cranial dural arteriovenous fistulas with hydrogel coated coils. *Interv Neuroradiol*, 2006. 12(4): p. 319-26.
128. Moenninghoff, C., et al., Outcomes After Onyx Embolization as Primary Treatment for Cranial Dural Arteriovenous Fistula in the Past Decade. *Acad Radiol*, 2020. 27(6): p. e123-e131.
129. Li, Y., et al., Onyx embolization for dural arteriovenous fistulas: a multi-institutional study. *J Neurointerv Surg*, 2022. 14(1).
130. Choo, D.M. and J.J. Shankar, Onyx versus nBCA and coils in the treatment of intracranial dural arteriovenous fistulas. *Interv Neuroradiol*, 2016. 22(2): p. 212-6.
131. McConnell, K.A., et al., Neuroendovascular management of dural arteriovenous malformations. *Neurosurg Clin N Am*, 2009. 20(4): p. 431-9.
132. Olivecrona, H. and J. Riives, Arteriovenous aneurysms of the brain, their diagnosis and treatment. *Arch Neurol Psychiatry*, 1948. 59(5): p. 567-602.
133. Sundt, T.M., Jr. and D.G. Piepgras, The surgical approach to arteriovenous malformations of the lateral and sigmoid dural sinuses. *J Neurosurg*, 1983. 59(1): p. 32-9.
134. Thompson, B.G., J.L. Doppman, and E.H. Oldfield, Treatment of cranial dural arteriovenous fistulae by interruption of leptomeningeal venous drainage. *J Neurosurg*, 1994. 80(4): p. 617-23.
135. Pradilla, G., et al., Surgical treatment of cranial arteriovenous malformations and dural arteriovenous fistulas. *Neurosurg Clin N Am*, 2012. 23(1): p. 105-22.
136. Steiger, H.J., D. Hänggi, and R. Schmid-Elsaesser, Cranial and spinal dural arteriovenous malformations and fistulas: an update. *Acta Neurochir Suppl*, 2005. 94: p. 115-22.
137. Hugosson, R. and K. Bergström, Surgical treatment of dural arteriovenous malformation in the region of the sigmoid sinus. *J Neurol Neurosurg Psychiatry*, 1974. 37(1): p. 97-101.
138. Lucas, C.P., et al., Sinus skeletonization: a treatment for dural arteriovenous malformations of the tentorial apex. Report of two cases. *J Neurosurg*, 1996. 84(3): p. 514-7.
139. D'Aliberti, G., G. Talamonti, and M. Collice, Sinus skeletonization. *J Neurosurg*, 1996. 85(4): p. 738-40.
140. Oran, I., M. Parildar, and A. Derbent, Treatment of slow-flow (type I) perimedullary spinal arteriovenous fistulas with special reference to embolization. *AJNR Am J Neuroradiol*, 2005. 26(10): p. 2582-6.
141. Narvid, J., et al., Spinal dural arteriovenous fistulae: clinical features and long-term results. *Neurosurgery*, 2008. 62(1): p. 159-66; discussion 166-7.
142. Krings, T. and S. Geibprasert, Spinal dural arteriovenous fistulas. *AJNR Am J Neuroradiol*, 2009. 30(4): p. 639-48.
143. O'Reilly, S.T., et al., Recognition of the variant type of spinal dural arteriovenous fistula: a rare but important consideration. *J Neurosurg Spine*, 2022: p. 1-5.
144. Krings, T., et al., Imaging in spinal vascular disease. *Neuroimaging Clin N Am*, 2007. 17(1): p. 57-72.
145. Ferch, R.D., M.K. Morgan, and W.R. Sears, Spinal arteriovenous malformations: a review with case illustrations. *J Clin Neurosci*, 2001. 8(4): p. 299-304.
146. Nishio, A., et al., Atypical spinal dural arteriovenous fistula with supply from the lateral sacral artery. *J Clin Neurosci*, 2007. 14(1): p. 65-8.
147. Nogueira, R.G., et al., Onyx embolization for the treatment of spinal dural arteriovenous fistulae: initial experience with long-term follow-up. Technical case report. *Neurosurgery*, 2009. 64(1): p. E197-8; discussion E198.

148. Krings, T., et al., Endovascular management of spinal vascular malformations. *Neurosurg Rev*, 2010. 33(1): p. 1-9.
149. Antonietti, L., et al., Long-term outcome in the repair of spinal cord perimedullary arteriovenous fistulas. *AJNR Am J Neuroradiol*, 2010. 31(10): p. 1824-30.
150. Rashad, S., et al., Management of spinal dural arterio-venous fistulas. Report of 12 cases and review of literature. *Clin Neurol Neurosurg*, 2014. 125: p. 81-6.
151. Saladino, A., et al., Surgical treatment of spinal dural arteriovenous fistulae: a consecutive series of 154 patients. *Neurosurgery*, 2010. 67(5): p. 1350-7; discussion 1357-8.
152. Rodesch, G. and P. Lasjaunias, Spinal cord arteriovenous shunts: from imaging to management. *Eur J Radiol*, 2003. 46(3): p. 221-32.
153. Song, D., et al., Spinal cord vascular malformations in children. *Neurosurg Clin N Am*, 2010. 21(3): p. 503-10.
154. Oldfield, E.H. Surgical treatment of spinal dural arteriovenous fistulas. in *Seminars in Cerebrovascular Diseases and Stroke*. 2002. Elsevier.
155. Jahan, R. and F. Vinuela. Vascular anatomy, pathophysiology, and classification of vascular malformations of the spinal cord. in *Seminars in Cerebrovascular Diseases and Stroke*. 2002. Elsevier.
156. Liu, A.-H., P. Gobin, and H. Riina, Endovascular surgery for vascular malformations of the spinal cord. *Operative Techniques in Neurosurgery*, 2003. 6(3): p. 163-170.
157. Jackson, J. and S. Partovi, Imaging of spinal cord vascular malformations. *Operative Techniques in Neurosurgery*, 2003. 6(3): p. 125-140.
158. Kim, L.J. and R.F. Spetzler, Classification and surgical management of spinal arteriovenous lesions: arteriovenous fistulae and arteriovenous malformations. *Neurosurgery*, 2006. 59(suppl_5): p. S3-195-S3-201.
159. Oldfield, E.H., et al., Successful management of spinal dural arteriovenous fistulas undetected by arteriography. Report of three cases. *J Neurosurg*, 2002. 96(2 Suppl): p. 220-9.
160. Steinmetz, M.P., et al., Outcome after the treatment of spinal dural arteriovenous fistulae: a contemporary single-institution series and meta-analysis. *Neurosurgery*, 2004. 55(1): p. 77-88.
161. Takai, K., T. Komori, and M. Taniguchi, Microvascular anatomy of spinal dural arteriovenous fistulas: arteriovenous connections and their relationships with the dura mater. *Journal of Neurosurgery: Spine*, 2015. 23(4): p. 526-533.
162. Bakker, N.A., et al., Recurrence rates after surgical or endovascular treatment of spinal dural arteriovenous fistulas: a meta-analysis. *Neurosurgery*, 2015. 77(1): p. 137-144.
163. Watanabe, Y., et al., Microsurgical or endovascular strategy for complete obliteration of spinal arteriovenous shunts in a single-institute 10-year retrospective study. *J Clin Neurosci*, 2020. 80: p. 195-202.
164. Fiaschi, P., et al., Spinal Dural Arteriovenous Fistulas: Clinical Results and Quality of Life Assessment with Surgical Treatment as a Crucial Therapy. The Joint Experience of Two Centers. *World Neurosurg*, 2019. 122: p. e270-e278.